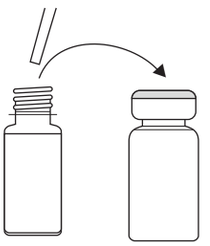
ABRAXIS® Glufosinate Derivatization Procedure
500060

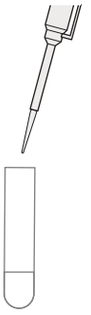
1. Derivatization Reagent Preparation

Reconstitute the derivatization reagent by adding 0.5 mL of the Derivatization Reagent Diluent (clear screw top glass vial) to the derivatization reagent vial (amber crimp top glass vial). Vortex and set aside.



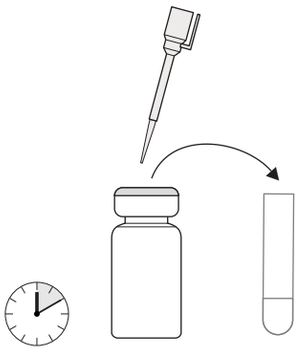
2. Addition of Standards & Samples to Test Tubes

Add 1 mL of each standard and sample to the appropriate labeled glass test tube.



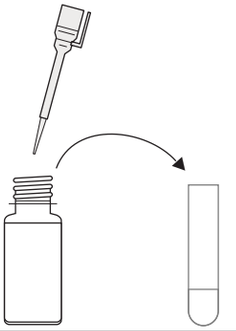
3. Addition of Derivatization Reagent

Add 10 uL of the reconstituted derivatization reagent (prepared in step 1) to each standard and sample glass test tube using a micropipette. Vortex each tube immediately after the addition of derivatization reagent for 15-30 seconds. Incubate tubes at room temperature for 15 minutes.



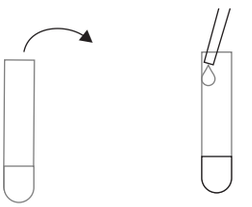
4. Addition of Buffer

Add 900 uL of Glufosinate Post-Derivatization Diluent to a new glass test tube for each standard and sample.



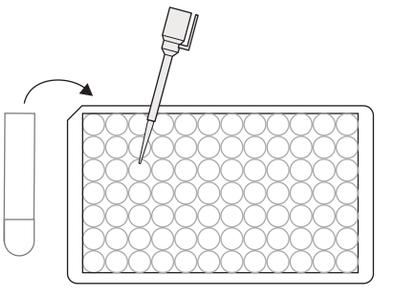
5. Dilution of Derivatized Standards & Samples to Test Tubes

Add 100 uL of derivatized standards and samples prepared in step 3 to each the Glufosinate Post-Derivatization Diluent glass test tube for each standard and samples prepared in step 4. Vortex each tube.



6. Analysis by ELISA

The derivatized and diluted standards and samples can then be analyzed using the Glufosinate ELISA Kit.



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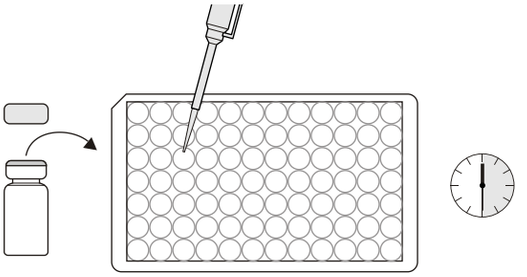
Version: 01

ABRAXIS® Glufosinate ELISA Plate
500060

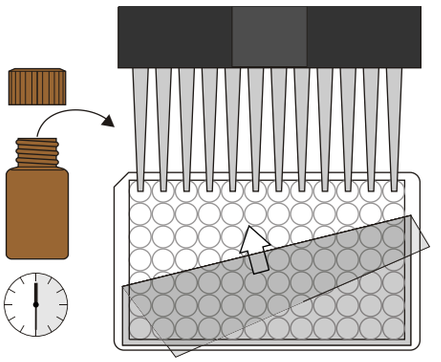
1. Addition of Standards, Samples

Add 100 uL of the derivatized and diluted standard solutions, control, or samples into the wells of the test strips according to the working scheme given. We recommend using duplicates or triplicates. Incubate for 30 minutes at room temperature.



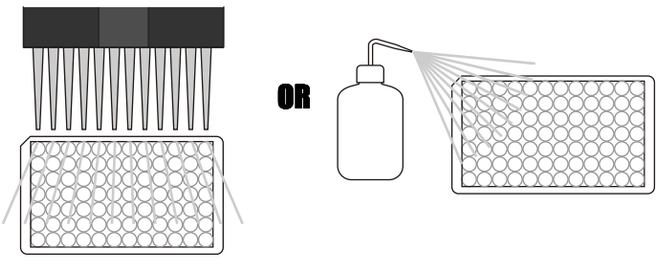
2. Addition of Reconstituted Enzyme Conjugate

Add 50 uL of the enzyme conjugate to the individual wells successively using a multi-channel pipette or a stepping pipette. Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a rapid circular motion on the benchtop for 30 seconds. Be careful not to spill contents.
Incubate for 30 minutes at room temperature.



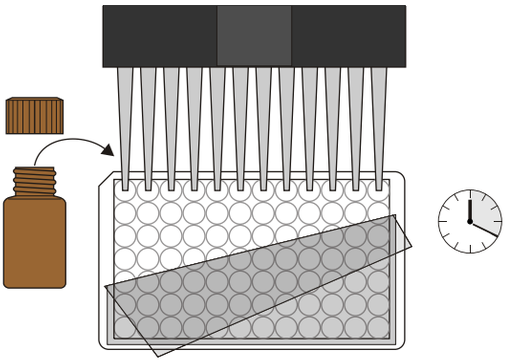
3. Washing of Plates

After incubation, remove the covering and vigorously shake the contents of the wells into a sink. Wash the strips three times with a multi-channel pipette or wash bottle using the 1X washing buffer solution. Please use at least a volume of 250 uL of washing buffer for each well and each washing step. Remaining buffer in the wells should be removed by patting the plate dry on a stack of paper towels.



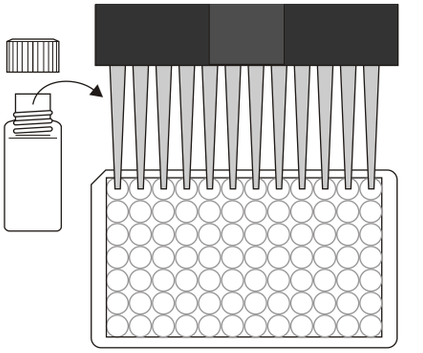
4. Addition of Substrate/Color Solution

Add 150 uL of substrate/color solution to the individual wells using a multi-channel pipette or a stepping pipette. Cover the wells with parafilm or tape and mix the contents by moving the strip holder in a rapid circular motion on the benchtop. Be careful not to spill contents.
Incubate the strips for 20 minutes at room temperature.



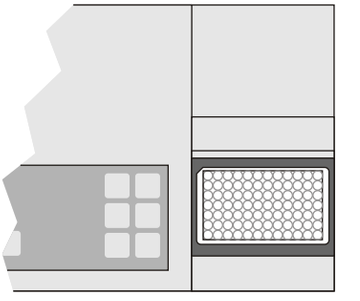
5. Addition of Stopping Solution

Add 100 uL of stop solution to the wells in the same sequence as for the substrate solution using a multi-channel pipette or a stepping pipette.



6. Measurement of Color

Read the absorbance at 450nm using a microplate ELISA reader. Calculate the results.



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