

ABRAXIS® Saxitoxins (PSP) Shipboard,

Accessory Pack

Product No. 530009

1. Intended Use

For the quantitative detection of Saxitoxin.

2. Safety Instructions

The standard solutions in the test kit contain small amounts of Saxitoxin. In addition, the substrate solution contains tetramethylbenzidine and the stop solution contains diluted sulfuric acid. Avoid contact of stop solution with skin and mucous membranes. If these reagents come in contact with the skin, wash with water.

3. Storage and Stability

The ABRAXIS[®] Saxitoxin (PSP) Shipboard Accessory Pack should be stored refrigerated (2 - 8°C). The solutions must be allowed to reach room temperature (20-25°C) before use. Reagents may be used until the last day of the month as indicated by the expiration date on the box. Consult state, local, and federal regulations for proper disposal of all reagents.

Working Instructions 4.

Materials Provided Α.

- 1. Diluent in dilution vials with blue stickers, 20, with labels (Dilution 1)
- Diluent in dilution vials with red stickers, 20, with labels (Dilution 2) 2.
- 4 mL glass vials with caps, 20, with labels (Sample Extract) 3.
- Pipette tips, 1 rack of 96, 10-200 µL 4
- 5. Plastic transfer pipettes, 20
- Microtiter plate frame with strip of blank wells (for zeroing reader) 6.
- Adhesive plate covers. 3 7.
- Simplified qualitative procedure/flow chart, data sheets (5), graph papers (5) 8.
- Β. Additional Materials (required but not provided with the test kit)
- ABRAXIS® Saxitoxin (PSP) Shipboard ELISA Microtiter Plate (PN 52255SB) 1.

Prior to Analysis C.

- Remove reagents from refrigerator and allow all reagents to warm to cabin temperature (20-30°C). 1.
- Prepare 1X wash buffer by emptying entire contents of Wash Buffer 5X Concentrate provided in the kit 2. into squeeze bottle. Fill to neck of bottle with DI water. Wash may be stored at room temperature up to a vear.
- 3. Extract each sample, transfer a portion (~1 mL) using plastic transfer pipette to 4 mL glass vials (w/black cap) if desired. Label vial appropriately. Extracted samples may be preserved by freezing on its side for up to 7 days.

D. After Extraction

- Dilution 1: Add 100 µL of extracted, filtered sample to a vial with blue stickered cap. Shake well and 1. label.
- 2. Dilution 2: Add 100 µL of Dilution 1 (blue stickered cap) to a vial with red stickered cap. Shake well and label. Dilution 2 vial is ready for analysis.
- Remove 2 strips from well rack located in silver pouch. Snap each strip into white frame, flush left. 3. Make sure to push the wells firmly into place.
- Note: 2 strip will accommodate 5 standards, 1 control, plus 2 samples, add another strip if analyzing more than 2 samples. Cover any unused wells with lab tape to prevent contamination.

- 4. Add 50 µL of each standard to the wells as follows; Std 0 to wells A1/B1, Std 1 to wells C1/D1, Std 2 to wells E1/F1, Std 3 to wells G1/H1, Std 4 to wells A2/B2, and Control to wells C2/D2. Add 50 µL of sample 1 (vial with red stickered cap) to wells E2 and F2. Continue adding 50 µL of further samples in duplicate. Add 50 µL of REAGENT 1 to each well in order of standard/sample addition.
- 5.
- Add 50 µL of REAGENT 2 to each well in order of standard/sample addition. 6.
- 7. Cover well plate with clear adhesive sheet provided. Mix by rotating in a circular motion on a flat surface for ~30 seconds. Set timer for 30 minutes incubation.

Note: Make sure plate is protected from direct sunlight.

- 8. After 30 min. incubation, remove adhesive plate cover, and empty wells by inverting plate into waste. Vigorously blot dry on a paper towel.
- 9. Fill wells to overflow with wash (Squeeze bottle). Invert plate and empty wells again into waste. Vigorously blot dry on a paper towel.
- 10. Repeat Step 9 three more times for a total of 4 washes.
- 11. Add 100 µL COLOR solution. Mix by rotation for ~30 seconds. Set timer for 30 minutes for incubation.

Note: Make sure plate is protected from direct sunlight.

- 12. After the 30 minute incubation. add 100 µL of STOP solution. Sample is ready to read.
- General Limited Warranty: Gold Standard Diagnostics warrants the products manufactured by the Company, against defects and workmanship when used in accordance with the applicable instructions for a period not to extend beyond the product's printed expiration date. Gold Standard Diagnostics makes no other warranty, expressed or implied. There is no warranty of merchantability or fitness for a particular purpose. This product is for research use only.

For ordering or technical assistance contact:

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Date these instructions are effective : 05/12/2025

Version: 02

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	PSP Ship Board Data Analysis Worksheet (Quantitative)
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Vessel	Kit Lot #	
Date	Kit Expiration	
Time	Technician	

	B/B ₀ , Result									ce 2	nth (01-12),	Longitude						
	Average Abs.									+Absorban	I-31), MM=mo	Long						Rev:111419
	Abs. A									$\frac{\text{Average} = \underline{\text{Absorbance 1} + \text{Absorbance 2}}{2}$	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surficlam	Latitude						Re
	Ð		(Sam 3)		(Sam 4)		(Sam 5)		(Sam 6)	Average =	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, YY=(10) T= Q for quahog, C for surfelam	Station #	01	02	03	6	05	
	Well	A3	B3	C3	D3	E3	F3	G3	Н3	Key:	How to label sar Where $VV=M$ YY=(10) T= Q	Station Log:			1		1	
	B/B ₀			>1.0?		B ₁ /B ₀ 83.2-95.8?		B ₂ /B ₀ 72.9-91.1?		B ₃ /B ₀ 58.2-78.0?		B ₄ /B ₀ 52.3-65.5?		Cone. 42- 78μg/100g?		B/B ₀ , Result		B/B ₀ , Result
(2)	Average Abs.		$B_{0}=$		$B_{l}=$		$B_{2}=$		$\mathrm{B}_{3^{=}}$		$B_{4}=$							
(>0.990?)	Abs.																	
$r^{2}=$	D	0	0	20	20	40	40	80	80	120	120	Control	Control		(Sam 1)		(Sam 2)	
I	Well	A1	Bl	CI	D1	E1	Fl	G1	H1	A2	B2	C2	D2	E2	F2	G2	H2 –	

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Kit Lot #	Kit Expiration	Technician	
Vessel	Date	Time	

	B/B ₀ , Result									ance 2	month (01-12),	Longitude						
	Average Abs.									1 +Absorb 2	(01-31), MM=	Γo						Rev:111419
	Abs.									$\frac{\text{Average} = \underline{\text{Absorbance 1} + \text{Absorbance 2}}{2}$	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), Y Y=(10) T= Q for quahog, C for surficlam	Latitude						
	D		(Sam 3)		(Sam 4)		(Sam 5)		(Sam 6)	Average	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, 1 YY=(10) T= Q for quahog, C for surfelam	Station #	01	02	03	04	05	
	Well	A3	B3	C3	D3	E3	F3	C3	H3	Key:	How to label sa Where VV=N YY=(10) T= (Station Log:						
	B/B ₀			>1.0?		B ₁ /B ₀ 83.2-95.8?		B_2/B_0 72.9-91.1?		B ₃ /B ₀ 58.2-78.0?		B ₄ /B ₀ 52.3-65.5?		Conc. 42- 78μg/100g?		B/B ₀ , Result		B/B ₀ , Result
(60	Average Abs.	B ₀ =		$B_{0}=$ $B_{1}=$			$B_{2}=$				$\mathrm{B}_{4^{=}}$							
(6000 02)	Abs.																	
2		0	0	20	20	40	40	80	80	120	120	Control	Control		(Sam 1)		(Sam 2)	
	Well	A1	Bl	C1	Dl	E1	Fl	G1	HI	A2	B2	C2	D2	E2	F2	G2	H2	

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t Analysis V	
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Date Kit Expiration Time Technician	Vessel	Kit Lot #	
	Date	Kit Expiration	
	Time	Technician	

		B/B ₀ , Result									ce 2	tth (01-12),	tude						
		Average Abs.									Average = $\frac{Absorbance 1 + Absorbance 2}{2}$	(01-31), MM=mor	Longitude						Rev:111419
		Abs.									- <u>Absorbance</u>	'YT number, DD=day (surfelam	Latitude						
		Ð		(Sam 3)		(Sam 4)		(Sam 5)		(Sam 6)	Average	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surficlam	Station #	01	02	03	04	05	
Technician		Well	A3	- B3	C3	D3	E3	F3	G3	H3	Key:	How to label sam Where VV=ML YY=(10) T= Q	Station Log:						_
		B/B ₀			>1.0?		B ₁ /B ₀ 83.2-95.8?		B ₂ /B ₀ 72.9-91.1?		B ₃ /B ₀ 58.2-78.0?		B4/B ₀ 52.3-65.5?		Conc. 42- 78μg/100g?		B/B ₀ , Result		B/B ₀ , Result
	(>0.990?)	Average Abs.				 B =		$B_{2}=$		B ₃ =		$B_{4^{\pm}}$				1		I	
Time	(>0)	Abs.																	
L	$r^{2=}$	Ð	0	0	20	20	40	40	80	80	120	120	Control	Control		(Sam 1)		(Sam 2)	
		Well	A1	B1	CI	DI	E1	F1	Gl	H1	A2	B2	C2	D2	E2	F2	G2	H2	

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	Average B/B ₀ , Abs. Result									Average = <u>Absorbance 1 + Absorbance 2</u> 2	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surfclam	Longitude						Rev:111419
	Abs.									= <u>Absorbance</u>	YYT 1 number, DD=day surfelam	Latitude						
	Ð		(Sam 3)		(Sam 4)		(Sam 5)		(Sam 6)	Average	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, YY=(10) T= Q for quahog, C for surfelam	Station #	01	02	03	04	05	
	Well	A3	B3	C3	D3	E3	F3	C3	H3	Key:	How to label sa Where $VV=M$ YY=(10) T= (Station Log:						
	B/B ₀			>1.0?		B ₁ /B ₀ 83.2-95.87		B ₂ / B ₀ 72.9-91.1?		B ₃ /B ₀ 58.2-78.0?		B4/B ₀ 52.3-65.5?		Conc. 42- 78μg/100g?		B/B ₀ , Result		B/B ₀ , Result
(206	Average Abs.		$\mathrm{B}_{0}=$		$\mathbf{B}_{I}=$		$\mathbf{B}_{2}=$	4	$\mathrm{B}_{3}=$		$\mathrm{B}_{4}=$							
(?0900<)	Abs.																	
$r^{2}=$	Ð	0	0	20	20	40	40	80	80	120	120	Control	Control		(Sam 1)		(Sam 2)	
	Well	A1	Bl	CI	D1	E1	FI	G1	H1	A2	B2	C2	D2	E2	F2	G2	H2	

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Kit Lot #	Kit Expiration	Technician
Vessel	Date	Time

-	ID Abs. Average B/B ₀ , Abs. Result		(Sam 3)		(Sam 4)		(Sam 5)		(Sam 6)	Average = $\underline{Absorbance 1 + Absorbance 2}$	How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surfclam	n Station # Latitude Longitude	01	02	03	04	50
	Well	A3	B3	C3	D3	E3	F3	B	H3	Key:	How to lab Where VV YY=(10)	Station Log:				<u> </u>	
-	B/B_0			>1.0?		B ₁ /B ₀ 83.2-95.8?		B ₂ /B ₀ 72.9-91.1?		B ₃ /B ₀ 58.2-78.0?		B ₄ /B ₀ 52.3-65.5?		Conc. 42- 78μg/100g?		B/B ₀ , Result	
(20	Average Abs.		$\mathrm{B}_{0}=$		$\mathbf{B}_{I}=$		B;=	70	$\mathrm{B}_{3}=$		$\mathrm{B}_{4=}$						
(?0900)	Abs.																
l	Ð	0	0	20	20	40	40	80	80	120	120	Control	Control		(Sam 1)		(Sam 2)
	Well	A1	Bl	CI	DI	E1	Fl	G1	HI	A2	B2	C2	D2	E2	F2	G2	H2









