ABRAXIS® Saxitoxins (PSP) Shipboard, Accessory Pack

Product No. 530009

1. Intended Use

For the quantitative detection of Saxitoxin.

2. Safety Instructions

The standard solutions in the test kit contain small amounts of Saxitoxin. In addition, the substrate solution contains tetramethylbenzidine and the stop solution contains diluted sulfuric acid. Avoid contact of stop solution with skin and mucous membranes. If these reagents come in contact with the skin, wash with water.

3. Storage and Stability

The ABRAXIS® Saxitoxin (PSP) Shipboard Accessory Pack should be stored refrigerated (2 - 8°C). The solutions must be allowed to reach room temperature (20-25°C) before use. Reagents may be used until the last day of the month as indicated by the expiration date on the box. Consult state, local, and federal regulations for proper disposal of all reagents.

4. Working Instructions

A. Materials Provided

1. Diluent in dilution vials with blue stickers, 20, with labels (Dilution 1)
2. Diluent in dilution vials with red stickers, 20, with labels (Dilution 2)
3. 4 mL glass vials with caps, 20, with labels (Sample Extract)
4. Pipette tips, 1 rack of 96, 10-200 µL
5. Plastic transfer pipettes, 20
6. Microtiter plate frame with strip of blank wells (for zeroing reader)
7. Adhesive plate covers, 3
8. Simplified qualitative procedure/flow chart, data sheets (5), graph papers (5)

B. Additional Materials (required but not provided with the test kit)

1. ABRAXIS® Saxitoxin (PSP) Shipboard ELISA Microtiter Plate (PN 52255SB)

C. Prior to Analysis

1. Remove reagents from refrigerator and allow all reagents to warm to cabin temperature (20-30°C).
2. Prepare 1X wash buffer by emptying entire contents of Wash Buffer 5X Concentrate provided in the kit into squeeze bottle. Fill to neck of bottle with DI water. Wash may be stored at room temperature up to a year.
3. Extract each sample, transfer a portion (~1 mL) using plastic transfer pipette to 4 mL glass vials (w/black cap) if desired. Label vial appropriately. Extracted samples may be preserved by freezing on its side for up to 7 days.

D. After Extraction

1. Dilution 1: **Add 100 µL of extracted, filtered sample to a vial with blue stickered cap**. Shake well and label.
2. Dilution 2: **Add 100 µL of Dilution 1 (blue stickered cap) to a vial with red stickered cap**. Shake well and label. Dilution 2 vial is ready for analysis.
3. **Remove 2 strips from well rack located in silver pouch. Snap each strip into white frame, flush left**. Make sure to push the wells firmly into place.

Note: 2 strip will accommodate 5 standards, 1 control, plus 2 samples, add another strip if analyzing more than 2 samples. Cover any unused wells with lab tape to prevent contamination.

4. **Add 50 µL of each standard to the wells** as follows; Std 0 to wells A1/B1, Std 1 to wells C1/D1, Std 2 to wells E1/F1, Std 3 to wells G1/H1, Std 4 to wells A2/B2, and Control to wells C2/D2. Add 50 µL of sample 1 (vial with red stickered cap) to wells E2 and F2. Continue adding 50 µL of further samples in duplicate.
5. **Add 50 µL of REAGENT 1** to each well in order of standard/sample addition.
6. **Add 50 µL of REAGENT 2** to each well in order of standard/sample addition.
7. **Cover well plate** with clear adhesive sheet provided. **Mix by rotating in a circular motion** on a flat surface for ~30 seconds. **Set timer for 30 minutes incubation**.

Note: Make sure plate is protected from direct sunlight.

8. After 30 min. incubation, **remove adhesive plate cover**, and **empty wells** by inverting plate into waste. Vigorously blot dry on a paper towel.
9. **Fill wells to overflow with wash** (Squeeze bottle). **Invert plate and empty wells again** into waste. Vigorously blot dry on a paper towel.
10. **Repeat Step 9 three more times** for a total of 4 washes.
11. **Add 100 µL COLOR solution**. Mix by rotation for ~30 seconds. **Set timer for 30 minutes for incubation.**

Note: Make sure plate is protected from direct sunlight.

12. After the 30 minute incubation, **add 100 µL of STOP solution**. Sample is ready to read.

For ordering or technical assistance contact:

Gold Standard Diagnostics
795 Horsham Road
Horsham, PA 19044
WEB: www.abraxiskits.com
Tel: (215) 357-3911
Ordering: info.abraxis@us.goldstandarddiagnostics.com
Technical Support: support.abraxis@us.goldstandarddiagnostics.com

Date these instructions are effective : 05/12/2025 Version: 02

GOLD STANDARD DIAGNOSTICS

PSP Ship Board Data Analysis Worksheet (Quantitative)

Vessel		Kit Lot #	
Date		Kit Expiration	
Time		Technician	

r^2=_____ (>0.990?)

Well	ID	Abs.	Average Abs.	B/B0	Well	ID	Abs.	Average Abs.	B/B0, Result
A1	0				A3				
B1	0		B0=		B3	(Sam 3)			
C1	20			>1.0?	C3				
D1	20		B1=		D3	(Sam 4)			
E1	40			B1/B0 83.2-95.8?	E3				
F1	40		B2=		F3	(Sam 5)			
G1	80			B2/B0 72.9-91.1?	G3				
H1	80		B3=		H3	(Sam 6)			
A2	120			B3/B0 58.2-78.0?	**Key:**	Average = Absorbance 1 +Absorbance 2 / 2			
B2	120		B4=		How to label samples: VVSSDDMMYYT Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surfclam				
C2	Control			B4/B0 52.3-65.5?	Station Log:	Station #	Latitude	Longitude	
D2	Control					01			
E2				Conc. 42-78µg/100g?		02			
F2	(Sam 1)					03			
G2				B/B0, Result		04			
H2	(Sam 2)					05			
				B/B0, Result					

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F2	(Sam 1)								
G2				B/B0, Result					
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B/B0, Result

Key: Average = (Absorbance 1 + Absorbance 2) / 2

How to label samples: VVSSDDMMYYT
Where VV=MD/SW, SS=Station number, DD=day (01-31), MM=month (01-12), YY=(10) T= Q for quahog, C for surfclam

Station Log:	Station #	Latitude	Longitude
	01		
	02		
	03		
	04		
	05		

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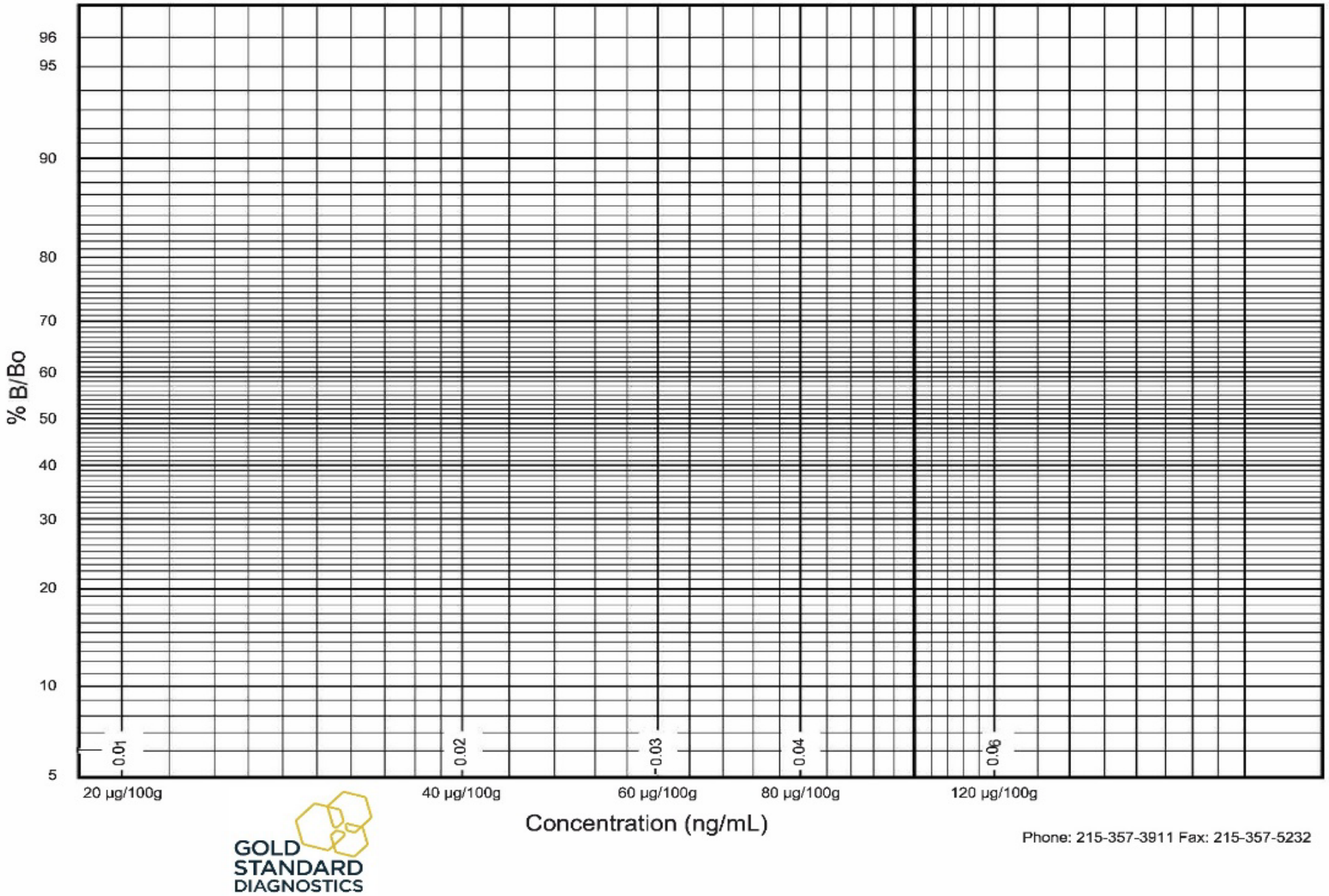
Saxitoxin SB



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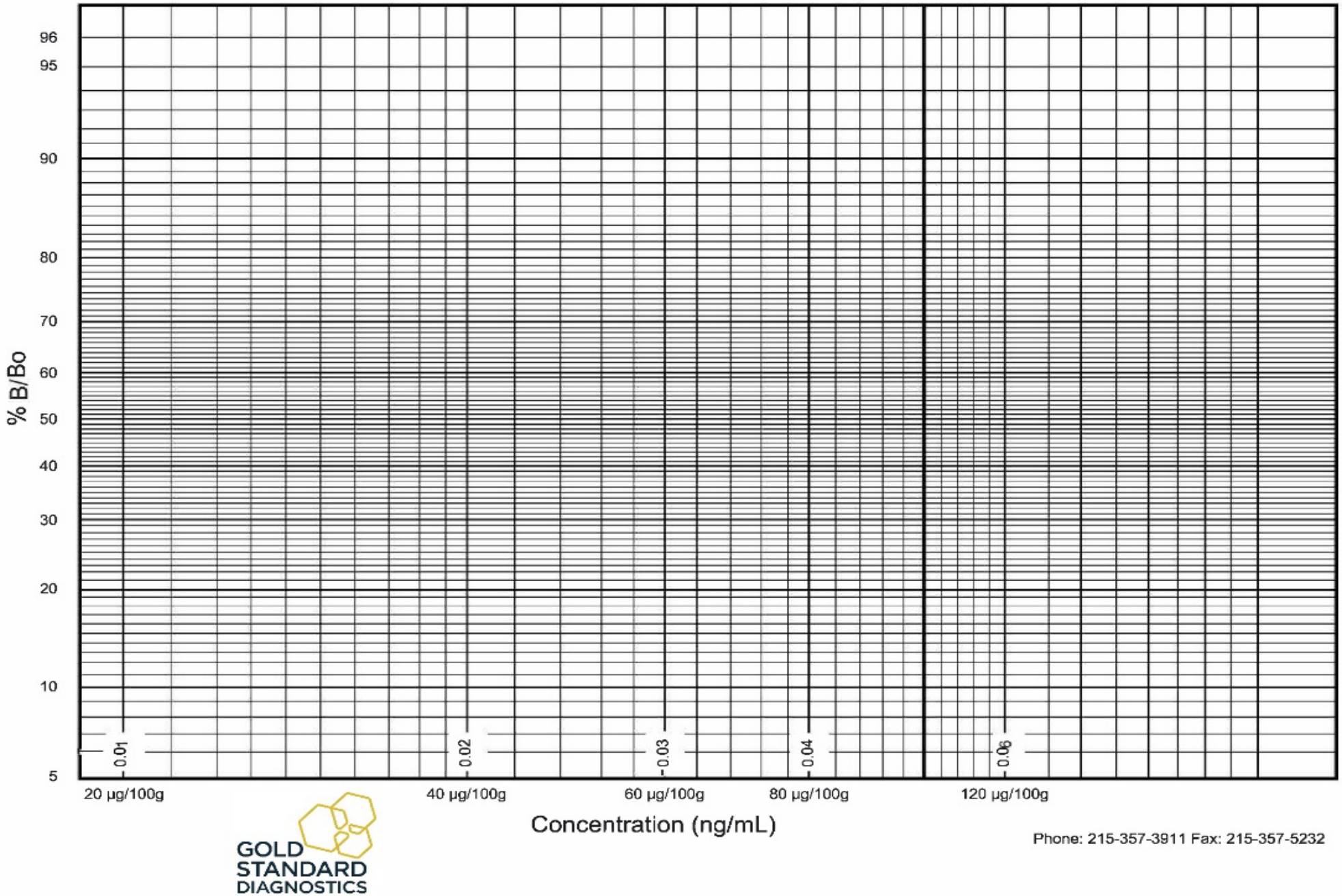
Saxitoxin SB



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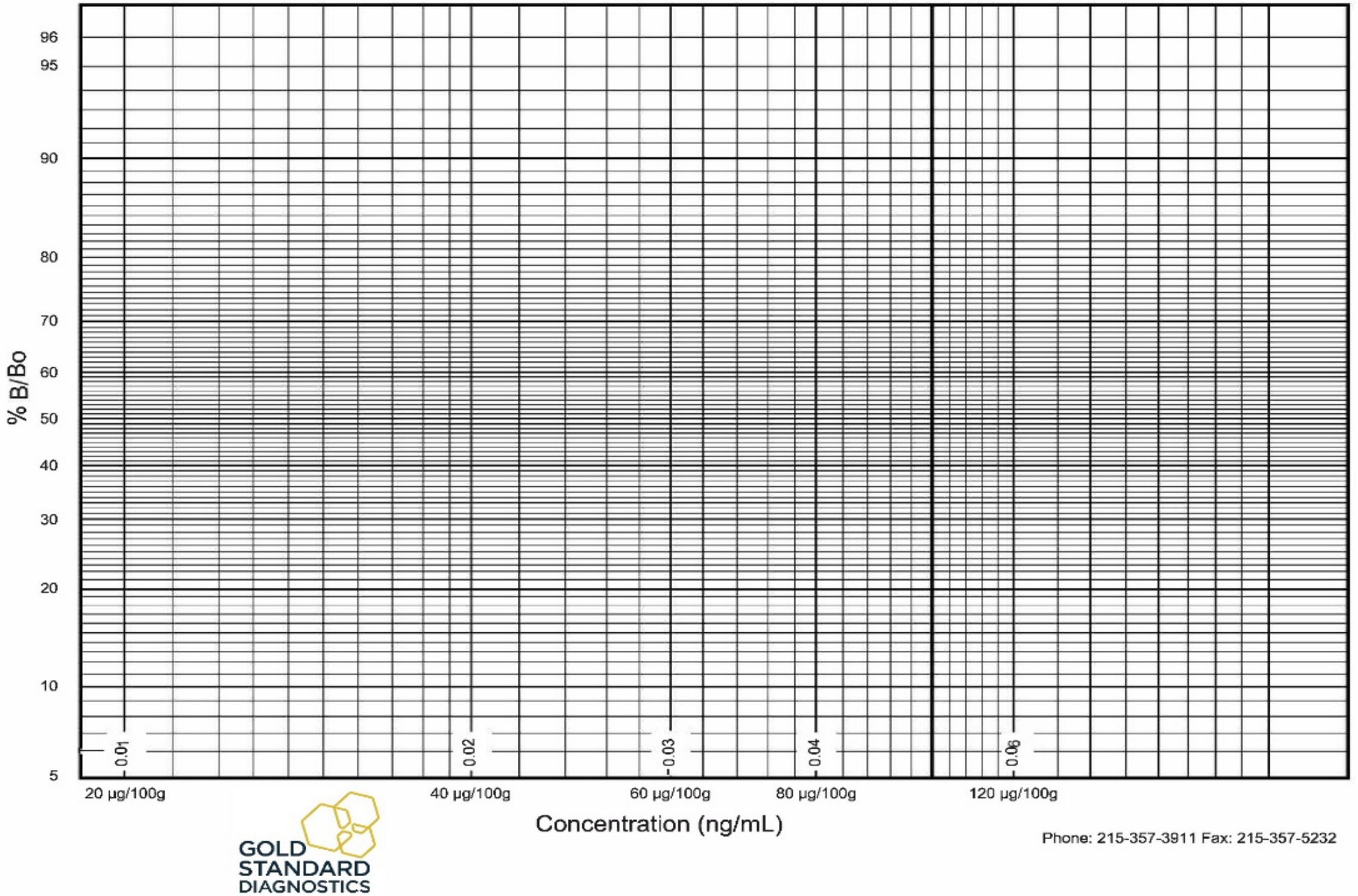

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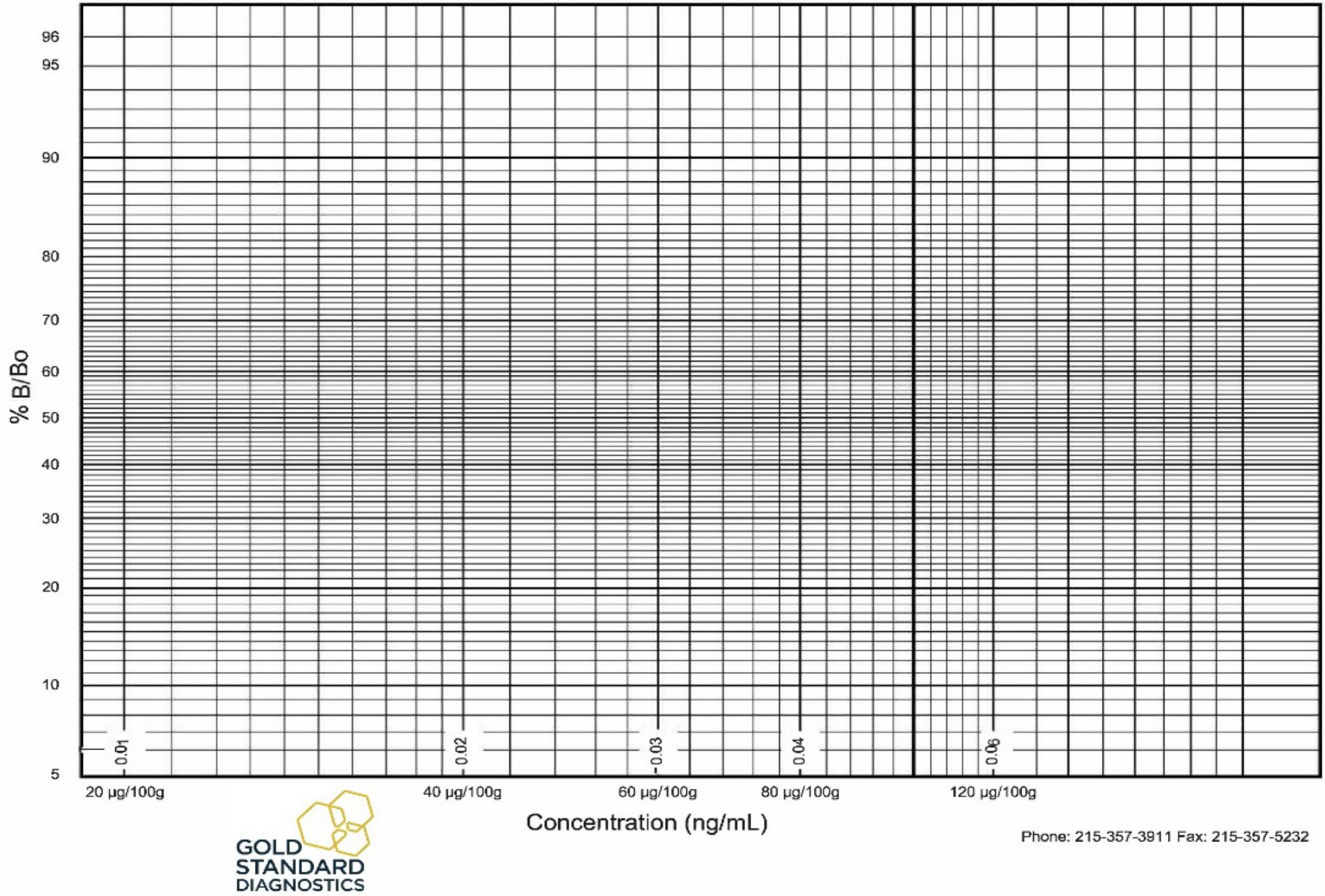

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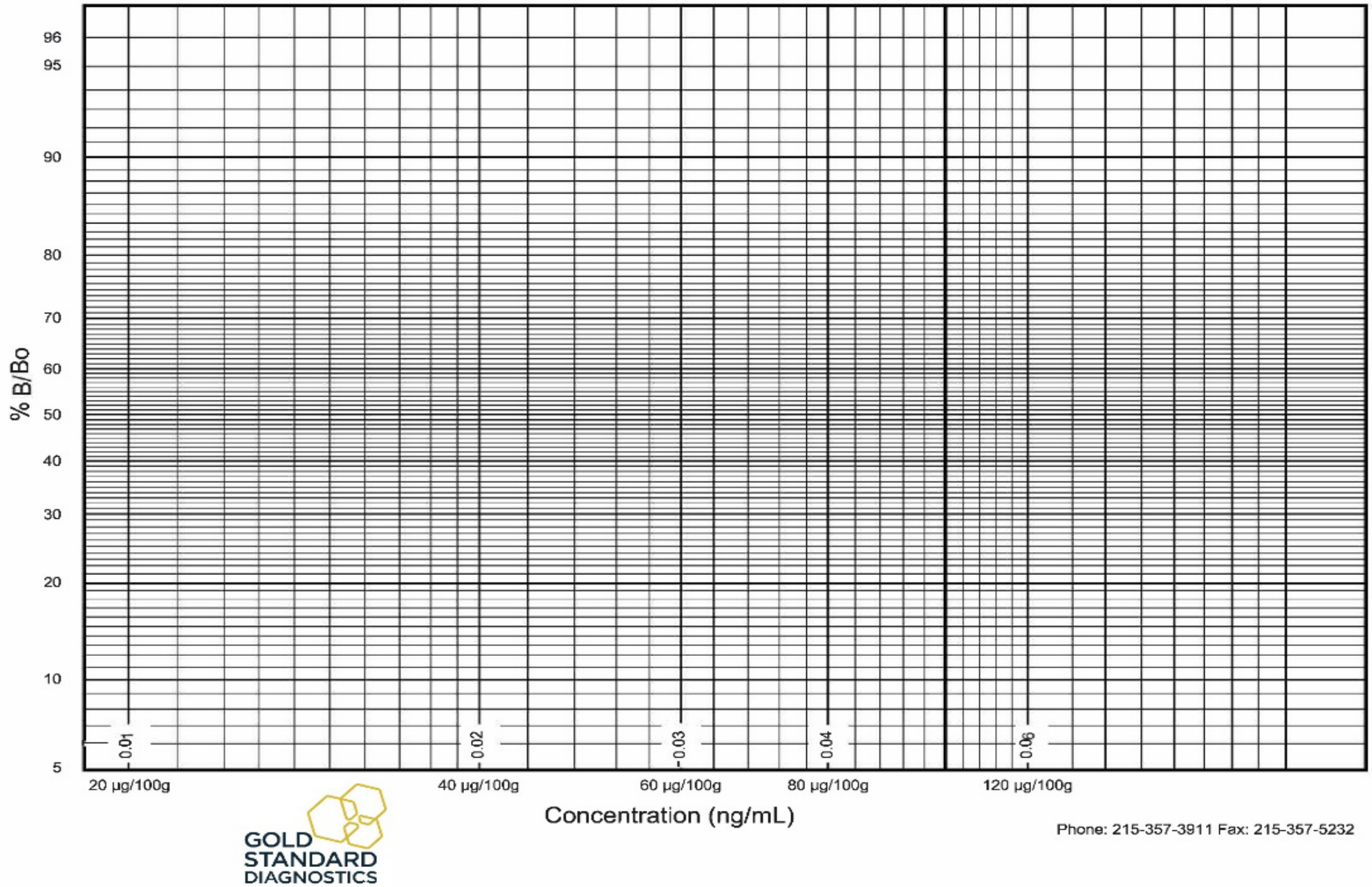

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